I. DNeasy protocol

A. Preparation of kit
   1. Add EtOH to AW1 and AW2 if needed
   2. Heat ATL and AL if they have precipitates
   3. Equipment
      a. 55C water bath
      b. 70C heat block

B. Sample digest
   1. put <25 mg tissue in 1.5 ml tube
   2. Add
      a. 180 ul buffer ATL
      b. 20 ul proteinase K
   3. Fin clips seem to take only 1 hour

C. Binding
   1. Vortex sample for 15s
   2. Add 200 ul buffer AL
      a. Mix again by vortex
      b. See white coagulant but it dissolves after heat
      c. Incubate at 70 C for 10 min
   3. Add 200 ul ethanol
      a. Mix by vortex
   4. Apply all to DNeasy spin column
      a. Spin 1 min >6000 g (9000 rpm in Eppendorf)
      b. Discard flow thru and tube
      c. Place in new tube

D. Washes
   1. Wash #1
      a. Apply 500 ul buffer AW1
      b. Spin 1 min, >6000g
      c. Discard flow thru and tube
      d. Place in new tube
   2. Wash #2
      a. Apply 500 ul buffer AW2
      b. Spin 2 min >20000g
      c. Discard flow thru and spin another minute
      d. Put column in clean 1.5 ml tube

E. Elution
   1. Elute #1
      a. Apply 100 ul buffer AE
      b. Incubate 1 min
      c. Spin 1 min in biorad; > 6000g (8500 rpm in biofuge)
   2. Elute #2
      a. Apply 100 ul buffer AE
      b. Incubate 1 min
      c. Spin 1 min in biorad; > 6000g (8500 rpm in biofuge)