#### Lecture 9: Mapping a gene defined by the mutation

I. Classical mapping II. Molecular mapping III. Positional cloning



Fig. 5.4

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P 
$$Q w^{+} m^{+}/w^{+} m^{+} \times O^{T} w m/Y$$
  
F<sub>1</sub>  $Q w^{+} m^{+}/w m \times w^{+} m^{+}/Y$   
F<sub>2</sub> males  
 $412 w^{+} m^{+}/Y$  Parental  $= \frac{412 + 389}{1192} \times 100 = 67.2\%$   
 $206 w^{+} m/Y$  Parental  $= \frac{206 + 185}{1192} \times 100 = 32.8\%$ 

Total 1192

Fig. 5.5



## Mapping with a CAPS marker



### A genetic map of DNA markers



#### **Physical map**



#### In Arabidopsis: 1 cM = about 200 kb (50 genes) In human 1cM = 1000 kb (~17 genes)

#### Yeast Artificial Chromosomes YAC

TEL	TRP1	ARS	CEN	Inserted foreign DNA		URA3	TEL
	Vector		insert size		Host		
	YAC: BAC: Cosmid: Lamda: Plasmid:		100-1000kb 80-300kb 20-50 kb 10-20kb		yeast bacterium bacterium bacterium		
			0.2-15kb		bacterium		

#### A MAP OF HUMAN CHROMOSOME 1



Figure 14–7. Linkage map of human chromosome 1, correlated with chromosome banding pattern. The histogram shows the distribution of all markers available for chromosome 1. Some markers are genes of known phenotype, but most are DNA markers based on neutral sequence variation. A linkage map, based on recombinant frequency analyses of the type described in this chapter, is in the center of the illustration. It shows only some of the markers available. Map distances are shown in centimorgans (cM, or m.u.). The total length of the chromosome 1 map is 356 cM; it is the longest human chromosome. The positions of some markers are cross-referenced to a diagram of subregions of chromosome 1 based on a standard banding pattern (such a diagram is called an idiogram). These kinds of correlations can be made only by using cytogenetic analysis (Chapter 17) and in situ hybridization. Most of the markers shown on the map are molecular, but several genes (highlighted in light green) also are included.

(Griffiths et al (Introduction to genetic 9 analyses)

## Steps in identifying the gene in a chromosome region

Step1: Categorize genes in the region Step2: Determine mRNA expression pattern EST Northern hybridization Step3: Determine changes between wt and mut Expression pattern DNA sequence Step4: Transgenic studies (SRY example) Case study: Cystic fibrosis CFTR (Cystic fibrosis transmembrane conductance regulator)

### How to determine where the genes are in a segment of DNA?

Fig. 11.19





3. Separate RNA samples by gel electrophoresis. Blot onto filter. Expose filter to labeled hybridization probe.



4. Wash away unhybridized probe. Make autoradiograph. 3 5 2 4 \_ \_ Tissue-specific \_ transcript \_ -(b) Over testis une titret 2.4 1.4 kb 0.2 Actin

#### **Testes-determining factor (TDF)**

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Fig. 11.21







Foreign DNA injected Injected eggs surgically Mice are born with foreign of newly fertilized egg. Injected eggs surgically implanted into uterus of "foster" mother and allowed to develop.



### **Cystic fibrosis: a case study**

Cystic fibrosis is an autosomal recessive disorder that results in defective transport of chloride ions through epithelial cells, and results from mutations in a gene, CFTR, which encodes a cAMP-regulated chloride channel. The primary expression of the defect is in the lungs: a sticky mucus secretion accumulates which is prone to chronic infections. Because there are no methods to culture lung cells routinely in the laboratory, in vivo gene therapy approaches have been adopted. As respiratory epithelial cells are differentiated, retroviral vectors cannot be used. Instead, gene therapy trials have used adenovirus vectors or liposomes to transfer a suitably sized CFTR minigene, either through a bronchoscope or through the nasal cavity.

#### Cloning cystic fibrosis transmembrane conductance regulator (CFTR)

Fig. 11.22a



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# (d)



#### CFTR



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