## Lecture 16: Functional Genomics II

- High-throughput genetic screens
- Insertional mutagenesis
  - Activation tagging
  - Enhancer trapping
- Modifier screens: enhancer and suppressor screens, synthetic lethal
- Yeast Two Hybrid Assay

# RNAi all genes on chromosome III -Gönczy, et al, 2000

- Goal: In *C. elegans*, determine function of all 2,300 genes on chromosome III
- RNAi constructs made for each gene
- Worms microinjected with doublestranded RNA
- Videos made of embryonic phenotypes



# **Phenotypes found in RNAi screen**



## Genome screen by feeding worms with dsRNA expressing *E. coli*



#### Genome-wide RNAi screens in Drosophila http://flyrnai.org

A functional genomic analysis of cell morphology using RNA-interference.

J Biol. 2003;2(4):27. Epub 2003 Oct 01

Kiger AA, Baum B, Jones S, Jones MR, Coulson A, Echeverri C, Perrimon N

**Genome-Wide RNAi Analysis of Growth and Viability in Drosophila Cells** 

Science. 2004 Feb 6;303(5659):832-5

Michael Boutros, Amy Kiger, Susan Armknecht, Kim Kerr, Marc Hild, Britta Koch, Stefan A. Haas, Heidelberg Fly Array Consortium, Renato Paro, Norbert Perrimon

#### Parallel Chemical Genetic and Genome-Wide RNAi Screens Identify Cytokinesis Inhibitors and Targets

PLoS Biol 2(12): e379.

- Ulrike S. Eggert, Amy Kiger, Constance Richter, Zachary E. Perlman, Norbert Perrimon, TimothyJ. Mitchison, Christine M. Field
- **Functional genomic analysis of the Wnt-Wingless Signaling Pathway**

Science. 2005 Apr 7

DasGupta R Kaykas A, Moon RT, Perrimon N

Terminal cytokinesis events uncovered after an RNAi screen.

Curr Biol. 2004 Sep 21;14(18):1685-93.

Echard A, Hickson GR, Foley E, O'Farrell PH.

## Genome-wide RNAi screens in mammalian cells

<u>A large-scale RNAi screen in human cells identifies new components of the p53</u> <u>pathway.</u>

Berns K, Hijmans EM, Mullenders J, Brummelkamp TR, Velds A, Heimerikx M, Kerkhoven RM, Madiredjo M, Nijkamp W, Weigelt B, Agami R, Ge W, Cavet G, Linsley PS, Beijersbergen RL, Bernards R. Nature. 2004 Mar 25;428(6981):431-7.

<u>An approach to genomewide screens of expressed small interfering RNAs in</u> <u>mammalian cells.</u>

Zheng L, Liu J, Batalov S, Zhou D, Orth A, Ding S, Schultz PG. Proc Natl Acad Sci U S A. 2004 Jan 6;101(1):135-40.

# Genome-wide screens in plants, planaria, ...

<u>RNA Interference Identifies a Calcium-Dependent Protein Kinase Involved in</u> <u>Medicago truncatula Root Development.</u>

Ivashuta S, Liu J, Liu J, Lohar DP, Haridas S, Bucciarelli B, VandenboschKA, Vance CP, Harrison MJ, Gantt JS.Plant Cell. 2005Nov;17(11):2911-21. Epub 2005 Sep 30.

**RNA silencing in plants.** 

Baulcombe D. Nature. 2004 Sep 16;431(7006):356-63. Review.

**Identification of genes needed for regeneration, stem cell function, and tissue homeostasis by systematic gene perturbation in planaria.** 

Reddien PW, Bermange AL, Murfitt KJ, Jennings JR, Sanchez Alvarado A.

Dev Cell. 2005 May;8(5):635-49.

Opening a new can of worms: a large-scale RNAi screen in planarians. 7 Newmark PA. Dev Cell. 2005 May;8(5):623-4.

## **Classical mutagenesis vs. RNAi**

- Diversity of mutations
  - (point mutations, deletions, inversions, etc.)
- Heritable, stable, and quantitative
- Saturating the genome requires hitting multiple genes repeatedly

- Give each gene equal attention
- High throughput
- Reasonably equivalent disruption of each locus
- "Mutations" automatically mapped
- Not heritable
- Doesn't generate full depletion of target RNA

# **Insertional mutagenesis for both forward and reverse genetics**

- Alternative to chromosome walking
  - To reduce time and effort required to identify mutant gene
  - Use inserted DNA to identify mutated gene
- Inserts randomly in chromosomes
- Conventional: DNA inserts disrupt genes
- Variations: activation tagging and enhancer trap

## **Insertional mutagens**

## • Transposable elements

- Mobile elements jump from introduced DNA
  - **P** elements in Drosophila
- Single-insertion elements
  - **T-DNA** in plants
    - Once insert, can't move again

# **Activation tagging**

- A variation on insertional mutagenesis
  - Makes gain-of-function mutations instead of loss-of-function mutations
  - An insertion that carries a strong constitutive promoter or enhancer
- Potential to identify gene function not detectable through loss-of-function screens
  - Useful for the following cases:
    - Functionally redundant genes
    - Genes required for viability

## Loss-of-function vs. gain-of-function usually give opposite phenotypes

## eyeless gene





Wild type



eyeless

Ectopic expression of eyeless 12

# **Activation tagging in** *Arabidopsis*

- Strong constitutive viral promoter
  - CaMV 35S
- Inserted randomly in genome – With T-DNA
- When inserts are near a gene promoter, the following results occur:
  - Activation
  - Constitutive expression
- Because many genes are expressed in specific cells or tissues, activation in all tissues can result in abnormal phenotypes



# Activation tagging examples in Arabidopsis

Identification of genes required for flowering time (A) and plant leaf and shape (B)

D. Weigel's lab



## Enhancer trapping in Arabidopsis

#### Minimal

promoter





# Enhancer trapping in Drosophila

- Use transposon *P* element
- Carries reporter gene -GFP
- Hops into genome
- When lands near enhancer, activates gene expression
- Expression similar to that of neighboring gene



# **Enhancer trapping in** *Drosophila*

- Reporter gene: Green Fluorescent Protein (GFP)
- The enhancer trap has inserted into a gene expressed in part of the fly eye



### Modifier screen example



*cal-1*: wild-type looking *ap1-1*: flower mutant *ap1-1 cal-1*: cauliflower



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#### Yeast Two Hybrid (Y2H) Assay

#### to test interaction between two proteins





#### LUFS



LUFS Q-rich (89-184, 449-470) 7 WD

**Q-rich (89-184, 449-470)** 7 WD

Use NCBI's Books to search for:

Yeast two hybrid assay Activation tagging Enhancer trap Etc.